

Allium negianum (Amaryllidaceae): a new species under subg. *Rhizirideum* from Uttarakhand Himalaya, India

Anjula Pandey¹, K. Madhav Rai², Pavan Kumar Malav¹, S. Rajkumar³

1 Division of Plant Exploration and Germplasm Collection, National Bureau of Plant Genetic Resources, New Delhi 110012, India **2** ICAR-National Bureau of Plant Genetic Resources, Regional Station Bhowali, Niglat 263132, Nainital, Uttarakhand, India **3** Division of Genomic Resources, ICAR-National Bureau of Plant Genetic Resources, New Delhi 110012, India

Corresponding authors: Anjula Pandey (anjuravinder@yahoo.com), Pavan Kumar Malav (pavan.malav@icar.gov.in)

Academic editor: Lorenzo Peruzzi | Received 3 March 2021 | Accepted 1 September 2021 | Published 15 October 2021

Citation: Pandey A, Rai KM, Malav PK, Rajkumar S (2021) *Allium negianum* (Amaryllidaceae): a new species under subg. *Rhizirideum* from Uttarakhand Himalaya, India. *PhytoKeys* 183: 77–93. <https://doi.org/10.3897/phytokeys.183.65433>

Abstract

A new species, *Allium negianum* (Amaryllidaceae), belongs to the genus *Allium* subg. *Rhizirideum*, sect. *Eduardia* is described here from the Uttarakhand Himalayan region of India. This taxon grows in Malari region of Niti valley in Chamoli district and Dharma valley of Pithoragarh district, Uttarakhand, India. It is a narrowly distributed species and morphologically more closer to *A. przewalskianum* Regel but differentiated by its tunic color of bulb, umbel with lax flowers, peduncle length, perigone colour, size and shape and leaf anatomy. Taxonomic delineation and relationship analysis based on nuclear ribosomal Internal Transcribed Spacers (ITS) region indicated that *A. negianum* is distinct and related to *A. przewalskianum*. This study provided a comprehensive description and comparison with *A. przewalskianum*, an identification key and notes on the distribution of the species.

Keywords

Allium negianum, India, *Rhizirideum*, Seasoning spice, Uttarakhand

Introduction

Allium L., one of the largest genera in the family Amaryllidaceae, has about 1,100 species distributed world-wide (Li et al. 2010; Govaerts et al. 2021). The genus *Allium* naturally occurs in dry seasons in the northern hemisphere and South Africa (Friesen et al. 2006; Nguyen et al. 2008; Neshati and Fritsch 2009). The primary centre of evolution for the ge-

nus extends across the Irano-Turanian bio-geographical region, and the Mediterranean basin and western North America are considered as the secondary centres of diversity (Friesen et al. 2006). The genus is characterized by bulbs that are enclosed within the membranous or fibrous tunics, free tepals, often a subgynobasic style and well-known characteristic plant odour and taste due to the presence of cysteine sulphoxides (Friesen et al. 2006). The classification of global species in the genus *Allium* is based on molecular phylogenetic analyses, which includes 15 subgenera and 56 sections (Friesen et al. 2006). The Indian *Allium* includes over 10 subgenera, 22 sections and 35–40 taxa excluding cultivated species distributed in different eco-geographical areas of the temperate and alpine regions of Himalayas sharing many taxa of Chinese origin (Pandey et al. 2008, 2017; Li et al. 2010). Indian Himalayan region has two distinct centres of diversity, the western Himalaya (over 85 per cent of total diversity) and the eastern Himalaya (6 per cent), covering the alpine-sub temperate region (2500–4500 m a.s.l.) (Gohil 1992; Pandey et al. 2008).

Globally *Allium* subg. *Rhizirideum* (G. Don ex Koch) Wendelbo s.str. has ca. 37 taxa that are included in four sections distributed mainly in Europe-East Asia, in China (Friesen et al. 2006; Choi et al. 2012; Jang et al. 2021) and also in Russia, Mongolia and Kazakstan (Sinityna et al. 2016; Friesen et al. 2020). *Allium senescens* L. of sect. *Rhizirideum* a species native to northern Europe and Asia from Siberia-Korea and also naturalized in parts of Europe, is an exception (Xu and Kamelin 2000; Li et al. 2010).

Taxa of the subg. *Rhizirideum* belong to the third and the most advanced evolutionary line, which is phylogenetically sister to taxa of the subg. *Allium* L., *Cepa* L., *Reticulatabulbosa* (Kamelin) N. Friesen and *Polyprason* Radic. (Friesen et al. 2006; Memariani et al. 2007; Li et al. 2010; Choi et al. 2012). The sect. *Eduardia* N. Friesen of the subg. *Rhizirideum* is mainly distributed in the western Himalaya with Pakistan on the west and Nepal and Tibet in the centre, and southwest China on the eastern side. Its habitat mainly comprises of mountainous, snow peak grassland, dry or rocky places in forests, subalpine meadows, steppes, sunny, saline areas, sandy deserts, stony and gravelly slopes, rocky crevices along the stream banks and damp places (Fritsch and Friesen 2002; Choi and Oh 2011; Choi et al. 2012).

Despite the importance of the genus *Allium* for the Indian region, meagre comprehensive studies have been attempted pertaining to molecular and taxonomic evaluation that led to gaps in the status of interspecific and infraspecific relationships among the taxa. Meagre taxonomic studies on the native taxa, unavailability of material for research, sporadic collections from under-explored/unexplored areas and lack of the published literature have led to the possibility of finding new taxonomic records from the Indian region (Pandey et al. 2008, 2017, 2021).

The subg. *Rhizirideum* is the smallest subgenus of *Allium* as per the flora of India, and it is represented only by the sect. *Eduardia* containing only one species, *A. przewalskianum* Regel. This taxon occurs in the scrub, drier slopes, ravines and rocky crevices (2000–4500 m a.s.l.) in Leh, Jammu and Kashmir and Spiti in Himachal Pradesh. The taxa under subg. *Rhizirideum* are characterized by the presence of several narrowly ovoid-cylindric bulbs, which borne on creeping rhizome usually covered with a common reticulate membrane, leaves shorter than scape, adaxially channeled and stamens slightly longer than perigone segments, spathe with a long beak, nearly 2 to 3 times longer than the base and hemispherical umbel. Most species share a basic chromosome

number of $x = 8$ and $2n = 16$ or 32 . Occurrence of a polyploid complex in different sections of the subgenus *Rhizirideum* indicated recent origin of taxa as supported by phylogenetic and biogeographical evidences (Li et al. 2010). Areas with geographical isolation are the driving force of underestimated speciation (Seregin et al. 2015).

A new taxon, *Allium negianum*, was collected from the Indo-Tibetan border area of Malari village, Niti valley of Chamoli district in Uttarakhand (India) in 2019 and identity was confirmed by the authors. It is distinct from its closest relative, *A. przewalskianum* Regel (Table 2), the only taxon of subg. *Rhizirideum*, sect. *Eduardia* in India. It is characterized by finely reticulated red-brown outer tunics, hemispherical umbel having lax flowers, spathe with a very long beak, deep purple tepals, asynchronous flowering and inner stamen filaments having longer and sharp teeth. In the present work, *A. negianum*, is described and illustrated here. Authors have examined the evidences from morphology, eco-geography, leaf anatomy, molecular study, and taxonomic delineation from other related species.

Materials and methods

Taxon sampling and morphological descriptor

A total of 110 plants representing 7 accessions of the new species were collected from the type locality and farmers' fields in the Niti region of Uttarakhand, India. For delimitation of the taxon with other related species, plants were grown in the Field Gene Bank (FGB) at the ICAR-National Bureau of Plant Genetic Resources (ICAR-NBPGR), Regional Station Bhowali (Nainital), Uttarakhand for comparative study of morphological characters. Data were recorded using the *Allium* descriptor with modifications from the published literature. The floral characters were measured with separate parts to the nearest ten points of the decimal. The seeds having uniform size and maturity were recorded for ultra-features of the characters using the Stereozoom Microscope (LMI, England, model no. SZM167), and the images were captured as JPEG. Ten replicate voucher herbarium specimens of the new species were prepared as per standard procedure and deposited in the National Herbarium of Cultivated Plants (code-NHCP) (Holotype) and CAL (Isotype).

The new species was compared with its closest relative using data derived from the study of specimens preserved in the herbaria of CAL, DD, E, K and NHCP and available literature. Due to its closer affinity with *A. przewalskianum*, all the specimens from diverse sources were critically examined. Taxonomic description and identification key were provided for *Allium negianum* and affined species.

Leaf anatomy

For leaf anatomy live plants were grown in the FGB at Regional Station Bhowali (Nainital), Uttarakhand. Leaf-blades were taken from a point 3–4 cm above the sheaths and fixed in 70% alcohol. Cross-sections were made at three different lengths of leaf and stained with Sartur solution (a mix of sudan III, aniline, chloral hydrate, lactic acid, iodine), the structure was studied, and analyzed with the help of a light

microscope (Olympus BH-2) and line diagrammes drawn. The outlines of cells were diagrammatically depicted (Fig. 1F).

Taxonomic delineation and relationship analysis

DNA extraction, amplification and sequencing

Genomic DNA of nine known species and one new taxon (Table 1) which was collected from western Himalayan region and maintained as live material at Field Gene Bank (FGB), ICAR- National Bureau of Plant Genetic Resources, Regional Station, Bhowali, was isolated from fresh leaves using spin column-based Qiamp DNA kit according to the suppliers' protocol. Selection of taxa for this study was mainly based on the fact that all taxa belong the third evolutionary line representing same eco-geographical areas, and were known by similar local names. This has resulted in confusion of their identity in Indian literature. The quantity and purity of the isolated genomic DNA was tested using the spectrophotometric method. The universal primers ITS1 and ITS4 (White et al. 1990) were used to amplify the ITS regions. The PCR protocol was run at 94 °C for 5 minutes; 30 cycles of 94 °C for 45 seconds, 55 °C for 45 seconds and 74 °C for 45 seconds and 74 °C for 5 minutes. PCR products were purified using Zymo DNA concentrator kit following the supplier's protocol. The purified PCR product was used in ABI 3730 DNA sequencer (Applied Biosystems) for generating sequences using PCR primers as sequencing primers. For remaining species from subgenus *Rhizirideum* and other related sub-genera, the ITS sequence were used from NCBI database.

Phylogenetic analysis based on the comparison of sequences

The generated DNA sequences from both the primers were checked for alignment using the BioEdit software. Multiple pairwise alignments of generated sequences and from NCBI database were made using ClustalW. The aligned sequences were used to generate the genetic distance between taxa and the evolutionary history, which was inferred by using the Maximum Likelihood method based on the Jukes-Cantor model using MEGA7.0 (Kumar et al. 2016).

Result and discussion

Taxonomic treatment

Allium negianum A.Pandey, K.M.Rai, Malav & S.Rajkumar, sp. nov.

urn:lsid:ipni.org:names:77220799-1

Figs 1, 2

Type. India, Uttarakhand: Chamoli, rocky areas (altitude 3000–4800 m), 22 Aug. 2019, KMR/AS/02/19 (Holotype: NHCP; Isotype: CAL; Seeds conserved in the National Genebank, New Delhi: IC258493).

Table 1. List of *Allium* taxa used to generate nuclear ITS sequence in the study.

S. no.	Taxon name	Subgenus	Section	NGB accession number	District; state
1	<i>Allium tuberosum</i> Rottdler ex Spreng.	<i>Butomissa</i>	<i>Butomissa</i>	IC353524	Almora; Uttarakhand
2	<i>Allium stracheyi</i> Baker	<i>Polyprason</i>	<i>Orioprasum</i>	IC567645	Pithoragarh; Uttarakhand
3	<i>Allium przewalskianum</i> Regel	<i>Rhizirideum</i>	<i>Eduardia</i>	IC632207	Leh; Jammu and Kashmir
4	<i>Allium negianum</i> sp. nov.	<i>Rhizirideum</i>	<i>Eduardia</i>	IC258493	Chamoli; Uttarakhand
5	<i>Allium sativum</i> L.	<i>Allium</i>	<i>Allium</i>	IC278243	Chamoli; Uttarakhand
6	<i>Allium ampeloprasum</i> var. <i>ampeloprasum</i>	<i>Allium</i>	<i>Allium</i>	IC353526	Pithoragarh; Uttarakhand
7	<i>Allium cepa</i> var. <i>cepa</i> L.	<i>Cepa</i>	<i>Cepa</i>	IC410711	Uttarkashi; Uttarakhand
8	<i>Allium cepa</i> L. var. <i>aggregatum</i> G.Don	<i>Cepa</i>	<i>Cepa</i>	AP/RP/2014	Chamoli; Uttarakhand
9	<i>Allium oschaninii</i> O.Fedtsch.	<i>Cepa</i>	<i>Cepa</i>	AP/2014	Voucher; Uttarakhand
10	<i>Allium schoenoprasum</i> L.	<i>Cepa</i>	<i>Schoenoprasum</i>	IC632213	Kargil; Jammu and Kashmir

Description. Herbs, hermaphrodite, 27–50 cm tall. Rhizome condensed, 6.5–8.5 mm long, oblique. Bulb clustered, cylindrical to narrowly ovoid, 0.8–1.2 cm in diameter, 6.8–12 cm long, outer tunic finely reticulate, reddish-dark brown, inner membranous, light-brick red. Leaves 4–6, slightly shorter than scape, 12–40 cm × 1.0–3.2 mm, erect, to semi-terete to terete, dark green; base slightly bulbous. Scape terete, semi-erect, covered with leaf sheaths at base only, stout, solid in cross-section (hollow in mature), 15–30 cm × 3.5–5.5 mm. Spathe 1-valved, persistent, beak very narrow-long, 2.5–4 mm. Inflorescence umbellate, hemispheric, 30–40 lax flowered. Peduncle subequal, 16–18 × 2–3 mm, without bulbils. Flowers bisexual, perigone campanulate, tepals dark purple with distinct green mid-line; inner tepals slightly longer than outer ones, oblong-lanceolate, apex acute, 6–8 × 3–4 mm; outer segments ovate to narrowly so, 5.5–6 × 2.5–3 mm. Stamens anthers oblong, yellow-purplish (on maturity), 2.3–2.6 mm long; filaments subequal, 6.8–8.5 mm, purple, slightly exerted, connate at base and adnate to perigone segments; outer ones subulate; inner ones broadened for 1/2–1/4 to their length, one sharp toothed on each side. Ovary sub-globose, purple-tinged, 3.6–4.8 × 1.8–3.5 mm. Style terete, exerted, stigma smooth, acute-acuminate, ovules 2 per locule. Capsules trigonous, 5–5.5 × 5.8–7.2 mm; seeds obovate with a prominent notch on one side, 3.2–4.0 × 1.9–1.9 mm, testa deep black. Plant has strong onion-garlic type aroma.

Habitat. Slopes, sandy soils along rivers and streams along the alpine meadows (altitude 3000–4800 m asl) in Sumna valley (villages Gamsali, Niti, Tolma, Kailashpur and Farkya) in Chamoli district near Malari glacier of India.

Etymology. The specific epithet, “*negianum*”, is named in honour of Late Dr. Kuldeep Singh Negi, an eminent explorer who has dedicated his life in collection of indigenous *Allium* species germplasm along with associated indigenous knowledge across the country. He was also instrumental in establishing the *Allium* Field Gene Bank (FGB) at the Regional Station, Bhowali, Uttarakhand. The entire germplasm of indigenous *Allium* species collected by him from remote areas of the country are characterized and successfully conserved at *Allium* FGB, Bhowali, Uttarakhand.

Vernacular/local name. Pharan, phran, jambu, sakua, sungdung, kacho, etc. (Pandey et al. 2021).

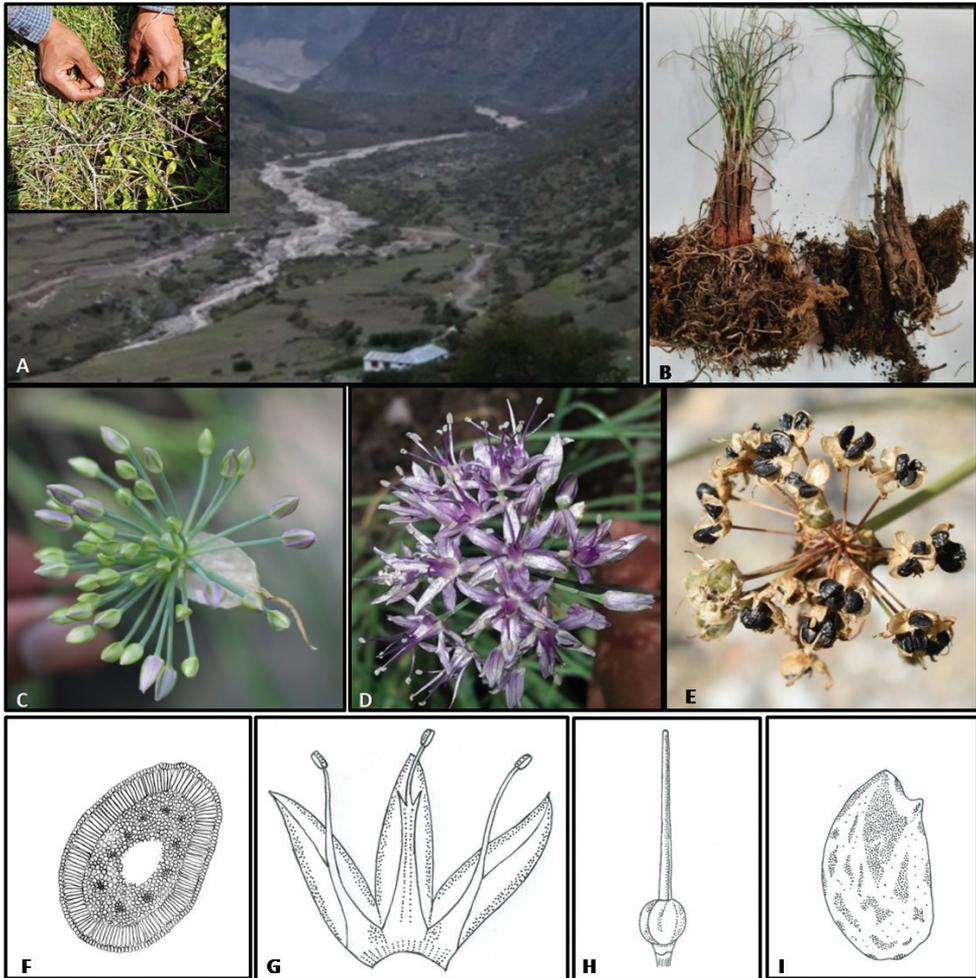


Figure 1. *Allium negianum* **A** general habitat **B** bulb covered with reticulate fiber on bulbs of *A. przewalskianum* (orange-red) and *A. negianum* (red-brown) **C** inflorescence and spathe with a very long beak, persistent **D** inflorescence **E** capsule with mature seeds **F** line-illustrations of transverse section of leaf showing hollow channel **G** longitudinal section of flower with stamen with two sharp teeth **H** ovary **I** seed with prominent beak (**C–I** magnification $\times 30$ – 40).

Phenology. Flowering and fruiting is from June to middle September (altitude 3000–4800 m a.s.l.).

Leaf anatomy. The transverse section of the leaf of *A. negianum* showed an elliptical outline. The epidermis has small cells covered with a thin cuticle layer, and stomata are narrowly distributed along the surface area. Single layered compactly arranged palisade tissue comprised of long cylindrical cells. The mesophyll cells are spongy tissue and compact in young leaf as well in the proximal ends of mature leaf while in the centre part of mature leaf, broken mesophyll cells are confused with fistulous leaf ap-

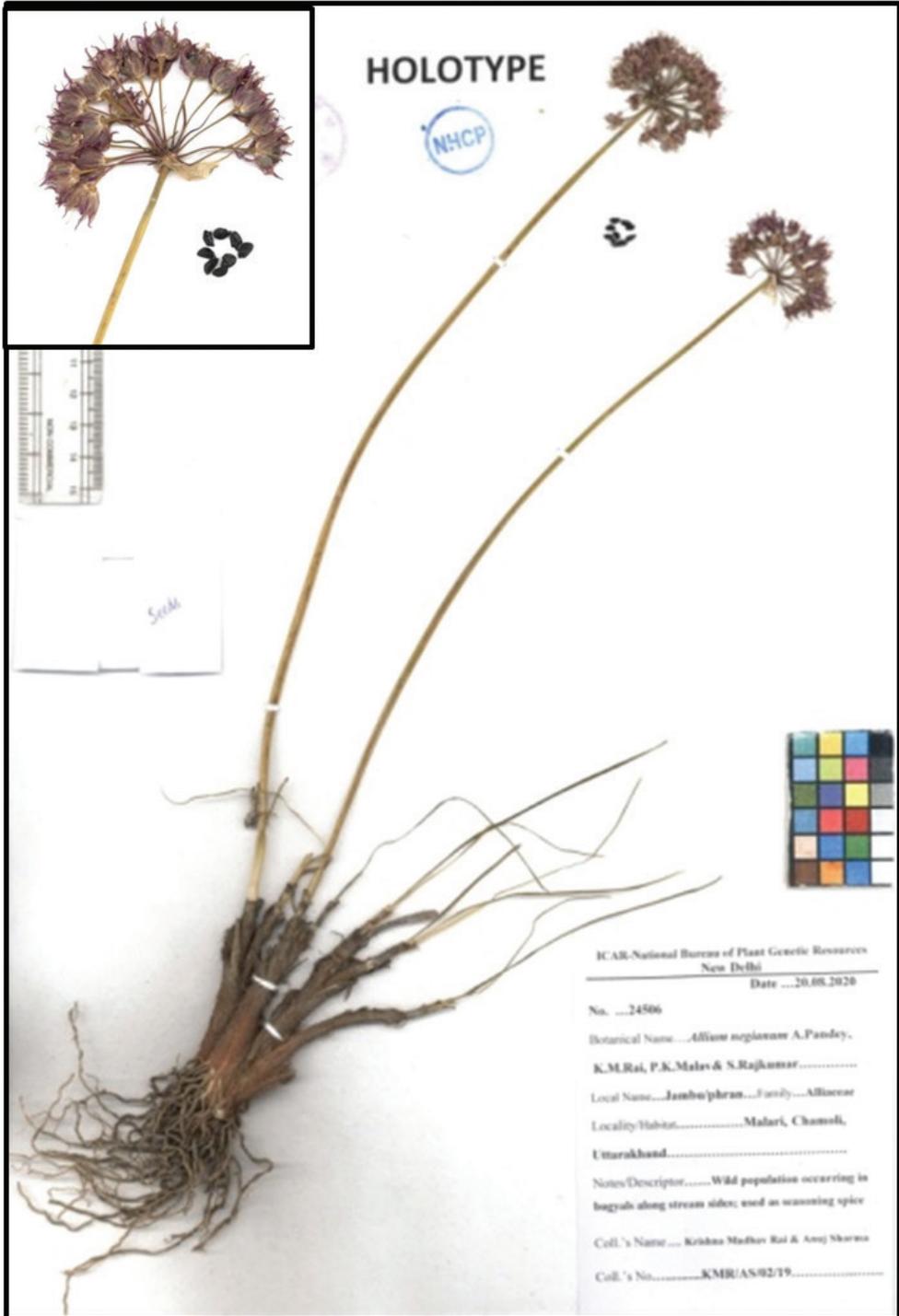


Figure 2. Holotype specimen of *Allium negianum* deposited in NHCP.

Table 2. Major morphological characters* (discriminating characters in **bold**) of *Allium negianum* in comparison with *A. przewalskianum*.

Character	<i>A. przewalskianum</i>	<i>A. negianum</i>
Habitat	Carbonaceous slates-gravel; 3300–5200 m	Grassy meadows, open sandy slopes, along rivers/ streams; 3000–4800 m
Plant habit	Erect	Semi-erect
Plant growth (under experimental condition)	Robust, shorter	Taller, plants and leaves
Plant height (cm)	20–45	27–50
Bulbs no. in cluster	2–4	2–7
Bulb no., shape	Cluster 3–4; cylindrical-narrowly ovoid	Cluster 4–8; cylindrical-narrowly ovoid
Bulb length (cm)	10.2–12.5	6.8–12
Bulb diameter (cm)	0.6–0.7	0.8–1.2
Tunic outer**	Finely reticulate; reddish-orange-brown	Finely reticulate; reddish-dark brown
Tunic inner	Membranous, brown-red	Membranous, orange-red
Rhizome type; size (mm)	Vertical, short; 3–5	Oblique; 7–12
Leaf no., colour	3–5, lighter brown-green	4–6, dark green
Leaf vs. scape	Much shorter than scape	Slightly shorter than scape
Leaf blade shape; apex	Linear, not fistular; obtuse to subrounded	Linear, filiform; acute
Leaf length (cm)	15–30	12–40
Leaf width (mm)	2.0–2.5	1–3.2
Leaf erectness	Erect	Erect-semierect
Leaf waxiness	Non-waxy	Waxy
Leaf cross section	Circular	Circular
Spathe valve if persistent	1(2)-valved, persistent	1-valved, persistent
Spathe valve shape, size	Ovate	Ovate-oblong
Spathe size (cm)	2–3 (two times the base; short, blunt)	4–6 (long narrow beak; 3 times the base)
Scape type	Solid, terete, erect, central	Solid, terete, erect to semi-erect, lateral-central
Scape size (cm)	30–40 × 0.2–0.35	20–50 × 0.36–0.48; 1/3-of the base
Pedicle vs. perigone	Subequal	2–3 times longer
Umbel flower opening pattern	Synchronous (80 per cent)	Asynchronous (30–40 per cent)
Umbel shape	Spherical-hemispherical, densely flowered, compact	Hemispherical, lax, loosely flowered
Umbel diameter (mm)	28.5–30.2	25.1–42.0
Umbel flower (no.)	25–40	30–40
Peduncle size (cm)	0.5–1.0	0.8–2.5
Flower size (cm)	0.4–0.5 × 0.3	0.5–0.5 × 0.4
Flower color	Pale red-purple pink (variable)	Dark purple (as recorded now)
Perigonium shape and color	Campanulate, pink-dark purple, tepal wide open	Campanulate, lilac, light to dark purple, tepal partly opened
Tepal shape	Ovate-lanceolate , apex obtuse	Elliptic, ovate-lanceolate ; apex-acuminate-mucronate
Tepal inner size length × width (cm)	0.3–0.4 × 0.2–0.3	0.5–0.6 × 0.3–0.4
Tepal outer size length × width (cm)	0.5–0.7 × 0.2–0.3	0.6–0.7 × 0.3–0.5
Tepal apex shape	Acute-acuminate	Acute, mucronate
Tepal maturity	Curved outwards	Slightly inwardly curved/rolled
Tepal mid-vein	Non-conspicuous; purple green-dark purple	Very conspicuous; green-light green
Anther length (mm)	6.1–9.3	6.8–8.5
Anther lobe length (mm)	Oblong-ovate, 1–2	Oblong, 1–2
Anther lobe color	Yellow-purple	Yellow-purple
Filament color	Yellowish-purple	Greenish yellow-purplish green
Filament length, position	Double the size of tepal; exerted,	Half the size of tepal; slightly exerted

Character	<i>A. przewalskianum</i>	<i>A. negianum</i>
Filament inner and outer anther	Inner – two sharp teeth up to 1/2 to 1/4 length of filament with broader base; outer narrower base	Inner – two shallower-sharper teeth up to 1/2 to 2/3 length of filament with base as wide as tepal; outer narrower base
Ovary shape	Ovoid – globose, wrinkled	Obovoid – subglobose
Ovary color	Purple green, tinged with purple	Dark-pale purple
Ovary style <i>vs.</i> anther (after pollination)	Much exerted , longer than the ovary	Slightly exerted or equal
Ovary stigma tip	Acuminate-acute	Acuminate
Stigma <i>vs.</i> stamen	Sub-equal	Slightly longer
Capsule shape	Ovoid	Sub-globose
Seed length (mm)	2.75–2.96	3.21–4.05
Seed width (mm)	1.55–1.59	1.92–1.97
Seed color	Dull black	Shiny black
Seed no./locule	2	2
1000 seed wt (g)	2.12	2.73
Odour when crushed#	Strong onion-light garlic	Strong onion-garlic

*data recorded from a minimum of 30 specimens of each taxon; additional 43 e-images; **: recorded immediately after uprooting; #: data on feedback; also refer Pandey et al. 2021.

pearance; 10–12 vascular bundles are arranged along with the palisade tissue across the entire circumference (Fig. 1F).

Seed morphology. Seed characters and testa sculptures represents a good taxonomic character in *Allium* (Neshati and Fritsch 2009; Celep et al. 2012; Lin and Tan 2017). Apparently, the seeds of the newly described species were marginally bigger than the related taxon, *A. przewalskianum*. Baasanmunkh et al. (2020) have discussed on the seed testa structure and its taxonomic implication for taxa of the subg. *Rhizirideum*. The seed size in *A. negianum* (Fig. 1I) measured 3.2–4.0 × 1.9–1.9 mm in contrast to 2.7–2.9 × 1.5–1.5 mm in the later taxon (Fig. 1I). The seeds of *A. negianum* are obovate in shape with a prominent notch on one side, gradually concave from edge to centre, with deep black and wrinkled testa.

The testa cell shape was irregularly hexagonal-pentagonal, loose with clear meshes of reticulated tissue. The anticlinal walls are usually raised, prominently small to intermediate granulate verrucae. The periclinal cells wall has several verrucae with irregular depressions. Study indicated that in subg. *Rhizirideum* testa cell shape varied from oval to irregular or oval to hemispherical; and seed length 1.30–2.35 mm, anticlinal wall were distinguished by nearly S type to straight and periclinal wall was flat to nearly convex with densely granulated verrucae (Baasanmunkh et al. 2020). *A. przewalskianum* was distinguished by irregular testa cells in a loose arrangement with reticulated tissue, straight to arched anticlinal walls, and concave periclinal walls with small to intermediate verrucae and granules (Lin and Tan 2017).

Distribution and ecology. The sect. *Eduardia* of the subg. *Rhizirideum* is distributed in the southern most range of the Himalayan region of India extending to China which is the centre of diversification. *Allium negianum* is a species recorded from the southernmost transitional zone between India and China. The distribution of *A. negianum* is restricted to the phytogeographical region of western Himalaya from Sumna

valley, Malari, Chamoli district of Uttarakhand, in western Himalaya, India where it commonly occurs along the open grassy meadows, sandy soils along rivers and streams occurring in the snow pasture lands along the alpine meadows (locally known as ‘bugyal’ or ‘bugial’) between 3000–4800 m a.s.l. (Fig. 1A; Fig. 3) in synanthropic habitats. It was reported growing as wild population in Darma valley of Pithoragarh, along Gori Ganga (also Gori Gad) river in the Munsiyari, Pithoragarh district, in Milam Glacier, in northeast of Nanda Devi, Uttarakhand, India. The seeds flowing with the melting snow led to its broader spread in the areas with good regeneration reported by the authors (Fig. 1A). Hence the taxon may be considered endemic in the area of study. Indiscriminate harvest of leaves and bulbs used for ‘seasoning’ purposes has threatened its wild population.

The first report on large scale cultivation of this taxon in Niti valley, Uttarakhand, as ‘seasoning allium spice’ called ‘jambu’ and ‘phran’ has been published (Pandey et al. 2021). Though the taxon was reported commonly under cultivation, the authors have observed the wild populations primarily from the above ‘type’ locality. The authors could not trace large scale cultivation of another taxon, *A. stracheyi* (used for same purpose and known by same local name) in the described locality in Uttarakhand (Pandey et al. 2021). Considering that *A. stracheyi* was a rare species reported from wild habitats in Uttarakhand Himalaya, the authors assume that the reports by Kuniyal and Negi (2018) on large scale cultivation may be referring to this newly described taxon which is also known by the same local name. Unfortunately, earlier studies on *A. stracheyi* did not provide any locality details, nor were the voucher specimens deposited in any herbaria of the material used in their study. Therefore, validation of the taxonomic identity could not be ascertained. Also, there is no occurrence record of the taxa belonging to subg. *Rhizirideum* from Uttarakhand, India.

Specimens examined (Paratypes). *Allium przewalskianum*: INDIA. Himachal Pradesh. Spiti, Takcha 25 Jul. 1972 U.C. Bhattacharya 48815(BSD); Tobo, Kinnaur, Lahul & Spity, 15 Sept. 2007, V.D. Verma & Ramchander (NHCP); Jammu & Kashmir. Ladakh, 25 July 1941, Ludlow & Sheriff8529 (BM); 8 Sep. 1941, Ludlow & Sheriff8571 (BM); Ganglas, 1 Aug. 1988, H.J. Chowdhury & B.P. Uniyal 86043 (BSD); 1880, Aitchinson376(CAL); Kashmir. Nubra, 24 July 1980, A.R. Naqshi & G.N. Dhar7370 under *A. stracheyi*; Leh (J&K), 8 Sept. 2014, K. Pradheep & P.S. Mehta1733 (NHCP); Leh (J&K), Nov. 2014, K. Pradheep HS21817(NHCP); Pangu lake, Luthum village, Leh (4500 m), s.s. Malik & D. Gautam15298 (NHCP); Uttarakhand. Malari, Chamoli, 10 Sept. 2019, Badal Singh & K. Madhav Rai HS24013 (NHCP); *Allium auriculatum*: Uttarakhand: Brahmmathya, district Chamoli, August 1988, K.S. Negi & M.N. Kopper 9387 (NHCP).

Online herbaria. *A. stoliczki*: Ladakh, Khaedubgla, 18 Aug. 1982, P.K. Hazra98623(K), 1985, Jacquemont V. Type (K); T. Thomson, Type (K); China, 1 Jan. 1872, Przewalski N.M., #s.n., Type (P); 01 Jan. 1884, Przewalski N.M., Type (P, K); 1872–1873, Przewalski N.M., #s.n., Type (G).

There are no records on the availability of this new taxon from Uttarakhand (Dasgupta 2006). Shah (2014) has raised doubts on reported cultivation of *A. przewalskianum*

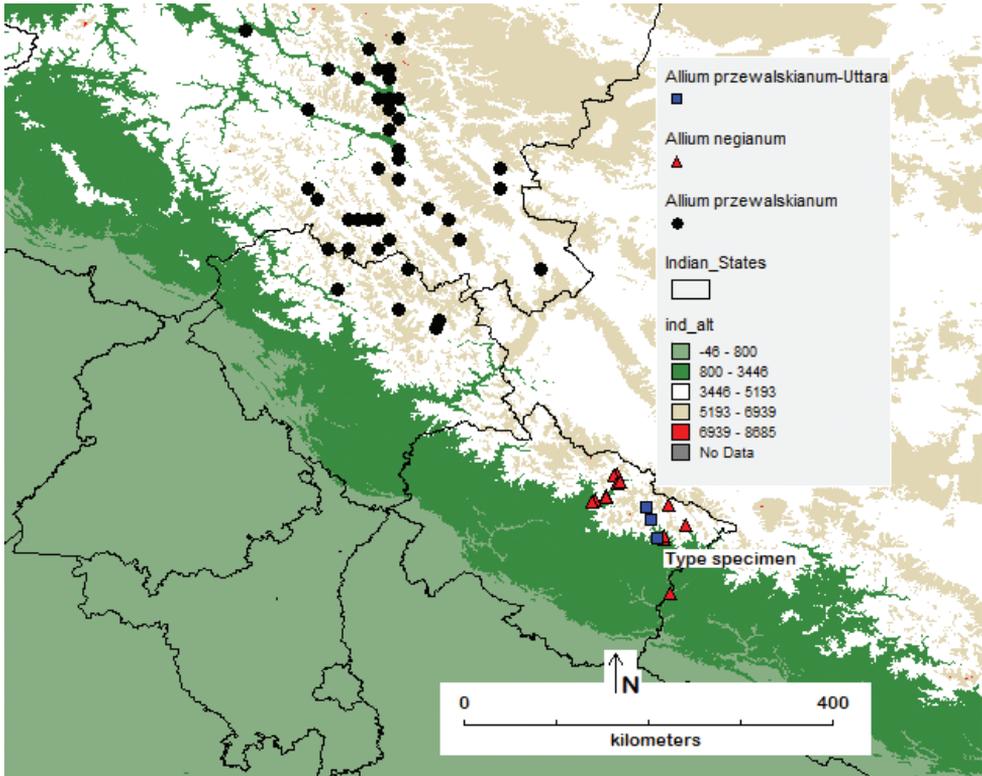


Figure 3. Distribution of taxa of *Allium* subg. *Rhizirideum* sect. *Eduardia* in India: *Allium negianum* and *A. przewalskianum* shown by the red triangle and black circled dots respectively; blue rectangle showed the occurrence of *A. przewalskianum* as per the data from GBIF (records of occurrence from Uttarakhand).

in Uttarakhand by Negi (2006). Also recorded data on the occurrence of allied taxon under *A. przewalskianum* from Gori, Kumaon, Uttarakhand (dated 16 June 2005) and Gori, Martoli, Uttarakhand (7 Oct. 2004) during the study of a total of 413 specimens in the GBIF database need critical study.

Note. *Allium negianum* was previously mistaken for identity as *A. stracheyi* as noted in the published records from India. Despite no morphological similarity with the latter taxon, Kuniyal and Negi (2018) referred 'phran' as *A. stracheyi*. In literature, it was also referred as *A. auriculatum* and *A. przewalskianum* due to morphological similarity of the outer tunics (Pandey et al. 2021). However, the present study demonstrated that *A. negianum* is clearly distinguished from *A. przewalskianum* and *A. stracheyi*, particularly characters of the bulb tunic color when fresh, umbel, teeth in filament and perigone size and color (Fig. 1B–H; Table 1). *Allium negianum* is diploid ($2n = 2x = 16$) (data not produced), whereas *A. przewalskianum* is reported to be tetraploid ($2n = 4x = 32$) as well as diploid with no stated morphological variation except the stout habit. Authors noted that *A. negianum* has robust plant habit, stronger plant aroma in wild habitat as compared to plants growing under cultivation. In contrast, the related taxon of the subg. *Rhizirideum*

is currently distributed in Jammu and Kashmir, Himachal Pradesh and adjoining parts in Nepal. *A. negianum* is reported from areas of Uttarakhand and only known from the type locality (altitude 3200–4800 m a.s.l.) and has never been collected from elsewhere in India and other parts of the world. Therefore *A. negianum* is said to be localized in distribution.

Upon critical examination of specimen of *A. auriculatum* deposited in the NHCP, all plant characters were found to be closer to *A. negianum*. Four specimens of this taxon were noted in label data as frequently growing on flat rocks in Brahmmathya, district Chamoli (3800 m asl.), Uttarakhand, used as leaves cooked as a vegetable.

Allium negianum is morphologically allied to a Chinese species *A. eduardi* Stearn that occurs on the dry slopes and plains in the adjoining regions of Mongolia and Russia and shares characters of spathe beak size, hemispherical umbel and perigone shape, but differs in having yellowish-brown bulb tunic color, tepal apex with a reflexed point and shorter stamen teeth length.

Taxonomic treatment

Two species, *A. przewalskianum* and *A. negianum*, of the subg. *Rhizirideum*, sect. *Eduardia* can be distinguished from *A. stracheyi* of the subg. *Polyprason* by using the following key.

Key to *Allium negianum* and related species

- 1 Bulbs cylindrical-narrowly oblong-ovoid, outer tunic fibrous, with finely reticulate texture, reddish-dark brown, leaves semiterete-terete **2**
- Bulbs cylindrical-narrowly ovoid, outer tunic fibrous scarious, brown-darkest brown, leaves narrow, fistulous..... ***A. stracheyi***
- 2 Bulbs outer tunic reticulate, reddish, inner tunic membranous, red-orange, rarely light brown; umbel compact globose, tepal pale-red to dark purple; filaments longer than perigone segments, inner ones broadened for 1/3–1/2 their length with shallow teeth; style very much exerted after anthesis ***A. przewalskianum***
- Bulbs outer tunic reticulate, reddish-brown, inner tunic membranous red; umbel hemi-spherical, lax; tepals dark purple-pink purple; filaments equal to perigone segments, inner ones broadened at the base for 2/3–1/3 of length, sharply marked teeth; style slightly exerted after anthesis ***A. negianum***

Taxonomic delineation and relationship analysis using nuclear ITS sequence

For taxonomic delineation and relationship analysis data set comprising 18 representative taxa from diverse subgenera were selected (Table 3; Fig. 4). The DNA sequence data set of nuclear Internal Transcribed Spacers (ITS) region used for phylogenetic analysis was generated for *Allium negianum* and other taxa used in the study. The generated ITS sequences and obtained ITS sequences from NCBI (Table 3) were used to construct the maximum likelihood tree. The tree with the highest log-likelihood is shown (Fig. 4). The percentage

of trees in which the associated taxa clustered together is shown next to the branches. Initial tree(s) for the heuristic search were obtained automatically by applying Neighbor-Join and BioNJ algorithms to a matrix of pairwise distances estimated using the Maximum Composite Likelihood (MCL) approach and selecting the topology with superior log likelihood value. The branch lengths measured in the number of substitutions per site.

Two major clades were found within *Allium*, comprising subgen. *Rhizirideum*, on one side and second cluster had four subg. *Butomissa*, *Allium*, *Polyprason* and *Cepa*. on the other side. This former group was divided in two sister clades, with first clade having *Allium przewalskianum*, *Allium negianum* sp. nov. *A. eduardii* (all from section *Eduardia*); *Allium subangulatum*, *A. polyrhizum* from sect. *Caespitosoprasum*; and *A. nutans*, *A. prostratum*, *A. spurium* and *A. spirale* in sect. *Rhizirideum*. One of the taxon *A. tenuissimum* from sect. *Tenuissima* grouped separately. Second clade was divided into subgenera, namely *Butomissa* with one taxon, *Allium tuberosum*; subg. *Allium*, with *Allium sativum* and *Allium ampeloprasum* var. *ampeloprasum*; subg. *Polyprason* having *Allium stracheyi*; subg. *Cepa* that was the largest having four taxa, *Allium cepa* var. *cepa*, *A. cepa* var. *aggregatum*, *A. oschaninii* and *A. schoenoprasum* from distinct sections.

Based on the likelihood tree, the new *Allium* taxon was observed to be closely related to *A. przewalskianum*, both of Indian Himalayan origin along with a Chinese taxon, *A. eduardii* to form distinct cluster supporting the morphological resemblance of this taxa with section *Eduardia* under subg. *Rhizirideum*. The species from other sections under same genus were distantly placed in the phylogenetic tree. The species which are found in same geographical area belong to different subgenera viz. *Allium*, *Cepa*, *Butomissa* and *Polyprason* were distantly placed and used as outgroup in determining the integrity of newly described species *Allium negianum*.

The above findings indicated that the new taxon is a distinct species and is closely related to *A. przewalskianum* and belongs to sect. *Eduardia* under subg. *Rhizirideum*. These findings supported the observations recorded using plant morphology, particularly the floral characters that were very distinct in both the taxa.

Recent advances in molecular phylogenetics have revolutionized our understanding of *Allium* taxonomy and evolution. However, the phylogenetic relationships in some *Allium* sections (such as the *Allium* sect. *Eduardia*) and the genetic bases of adaptative evolution remain poorly understood for the Indian taxa (Pandey et al. 2021). Molecular phylogeny study of the wild *Allium* in different centers of diversity (Nguyen et al. 2008; Xie et al. 2019; Jang et al. 2021) has helped in unlocking many aspects of the taxon relationships. The present study uncovered a new species relationship with its closest allied species and suggested that the selective habitat pressure has played an important role in the adaptation and evolution of *Allium* in this habitat which will facilitate uncover more taxa in the genus.

Conclusions

Allium negianum, a new species under the subg. *Rhizirideum*, is described using live and herbarium specimens. With the inclusion of this taxon, in the subg. *Rhizirideum* of the sect. *Eduardia* there are two taxa in India, and the latter one *A. negianum* was report-

Table 3. Details of nuclear ITS sequence used in present study.

Sl. No.	Species	Genbank accession number
1	<i>Allium tuberosum</i> Rottler ex Spreng.	MZ567234 (present study)
2	<i>Allium stracheyi</i> Baker	MZ567226 (present study)
3	<i>Allium przewalskianum</i> Regel	MZ567224 (present study)
4	<i>Allium negianum</i> sp. nov.	MZ567225 (present study)
5	<i>Allium sativum</i> L.	MZ567230 (present study)
6	<i>Allium ampeloprasum</i> L. var. <i>ampeloprasum</i>	MZ567231 (present study)
7	<i>Allium cepa</i> L. var. <i>cepa</i>	MZ567228 (present study)
8	<i>Allium cepa</i> L. var. <i>aggregatum</i> G.Don	MZ567232 (present study)
9	<i>Allium oschaninii</i> O.Fedtsch.	MZ567229 (present study)
10	<i>Allium schoenoprasum</i> L.	MZ567227 (present study)
11	<i>Allium eduardii</i> Stearn ex Airy Shaw	MK917745
12	<i>Allium subangulatum</i> Regel.	AJ411870
13	<i>Allium tenuissimum</i> L.	AJ411846
14	<i>Allium nutans</i> L.	JN864787
15	<i>Allium prostratum</i> Trevi.	LN867014
16	<i>Allium spurium</i> G.Don.	LN867017
17	<i>Allium spirale</i> Willd.	JN864784
18	<i>Allium polyrhizum</i> Turcz. ex Regel	MK917742

Source: S. no. 1–10 refer table 1; 11–18: NCBI

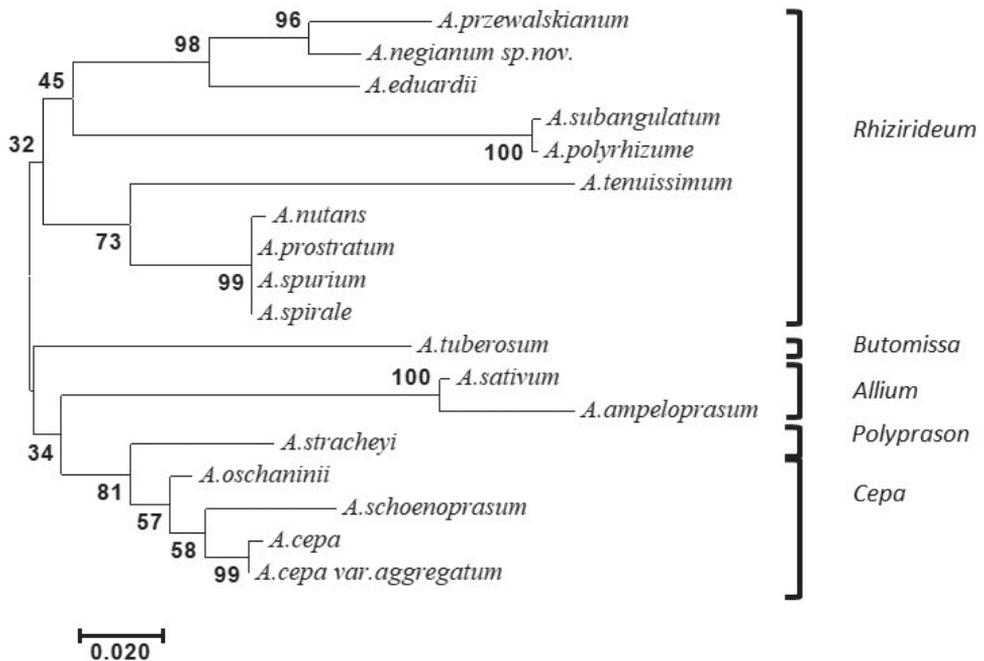


Figure 4. Maximum likelihood tree from nuclear ribosomal ITS sequence from *Allium* taxa showing distinctness of *Allium negianum* sp. nov.

edly restricted to the Uttarakhand flora. Samples of this taxon collected during earlier explorations that remained unidentified will be designated with this new name and conserved as seed in the National Gene Bank (NGB), New Delhi and vegetative material will be maintained in the Field Gene Bank (FGB) at Bhowali, Uttarakhand, India.

Acknowledgements

The authors are especially thankful to the Head, Division of Plant Exploration and Germplasm Collection and the Officer-in-charge, Regional Station Bhowali (Nainital), Uttarakhand to facilitate repeated surveys and field observations at the type locality under Global Environment Facility and National Exploration Plan (2019–20) programmes and for growing the germplasm. For the line diagrams help rendered by Mr. RK Pamarthi and Mr. Kunal Patil is greatly acknowledged.

References

- Baasanmunkh S, Lee JK, Jang JE, Park MS, Friesen N, Chung S, Choi HJ (2020) Seed morphology of *Allium* L. (Amaryllidaceae) from Central Asian Countries and its taxonomic implications. *Plants* 9(9): e1239. <https://doi.org/10.3390/plants9091239>
- Celep F, Koyuncu M, Fritsch RM, Kahraman A, Dogan M (2012) Taxonomic importance of seed morphology in *Allium* (Amaryllidaceae). *Systematic Botany* 37(4): 893–912. <https://doi.org/10.1600/036364412X656563>
- Choi HJ, Oh BU (2011) A partial revision of *Allium* (Amaryllidaceae) in Korea and north-eastern China. *Botanical Journal of the Linnean Society* 167(2): 153–211. <https://doi.org/10.1111/j.1095-8339.2011.01166.x>
- Choi HJ, Giussani LM, Jang CG, Oh BU, Cotañ-Sanchez JH (2012) Systematics of disjunct north-eastern Asian and northern North American *Allium* (Amaryllidaceae). *Botany* 90(6): 491–508. <https://doi.org/10.1139/b2012-031>
- Dasgupta S (2006) Alliaceae. In: Singh NP, Sanjappa M (Eds) *Fascicles of Flora of India*, no. 23. Botanical Survey of India, Calcutta, India, 18–19.
- Friesen N, Fritsch RM, Blattner FR (2006) Phylogeny and new intrageneric classification of *Allium* (Alliaceae) based on nuclear ribosomal DNA ITS sequences. *Aliso* 22: 372–395. <https://doi.org/10.5642/aliso.20062201.31>
- Friesen N, Vesselova P, Osmonaly B, Sitpayeva G, Luferov A, Shmakov A (2020) *Allium tok-sanbaicum* (Amaryllidaceae), a new species from Southeast Kazakhstan. *Phytotaxa* 494(3): 251–267. <https://doi.org/10.11646/phytotaxa.494.3.1>
- Fritsch RM, Friesen N (2002) Evolution, domestication and taxonomy. In: Rabinowitch HD, Currah L (Eds) *Allium crop science: recent advances*. CABI Publishing, Wallingford, 5–30. <https://doi.org/10.1079/9780851995106.0005>
- Gohil RN (1992) Himalayan representatives of *Alliums*. In: Hanelt P, Hammer K, Knupffer H (Eds) *The genus Allium: taxonomic problems and genetic resources*. Gatersleben, Germany, 335–340.
- Govaerts R, Kington S, Friesen N, Fritsch R, Snijman DA, Marcucci R, Silverstone-Sopkin PA, Brullo S (2021) World checklist of Amaryllidaceae. Facilitated by the Royal Botanic Gardens, Kew. <http://apps.kew.org/wcsp/> [accessed 21 May 2021]
- Jang JE, Park J-S, Jung J-Y, Kim D-K, Yang S, Choi HJ (2021) Notes on *Allium* section *Rhizirideum* (Amaryllidaceae) in South Korea and northeastern China: With a new species from Ulleungdo Island. *PhytoKeys* 176: 1–19. <https://doi.org/10.3897/phytokeys.176.63378>

- Kumar S, Stecher G, Tamura K (2016) MEGA7: Molecular evolutionary genetics analysis version 7.0 for bigger datasets. *Molecular Biology and Evolution* 33(7): 1870–1874. Epub-2016Mar22. <https://doi.org/10.1093/molbev/msw054>
- Kuniyal CP, Negi BS (2018) Cultivation of the Himalayan seasoning *Allium* in a remote village of Uttarakhand, India. *Journal of Threatened Taxa* 10(11): 12614–12617. <https://doi.org/10.11609/jott.3807.10.11.12614-12617>
- Li QQ, Zhou SD, He XJ, Yu Y, Zhang YC, Wei XQ (2010) Phylogeny and biogeography of *Allium* (Amaryllidaceae: Alliaceae) based on nuclear ribosomal internal transcribed spacer and chloroplast rps16 sequences, focusing on the inclusion of species endemic to China. *Annals of Botany* 106(5): 709–733. <https://doi.org/10.1093/aob/mcq177>
- Lin CY, Tan DY (2017) Seed testa micromorphology of thirty-eight species of *Allium* (Amaryllidaceae) from central Asia and its taxonomic implications. *Nordic Journal of Botany* 35(2): 189–200. <https://doi.org/10.1111/njb.01259>
- Memariani F, Joharchi MR, Khassanov FO (2007) *Allium* L. subgen. *Rhizirideum* sensu lato in Iran, two new records and a synopsis of taxonomy and phytogeography. *Iranian Journal of Botany* 13(1): 12–20. <https://www.sid.ir/en/journal/ViewPaper.aspx?id=101166>
- Negi KS (2006) *Allium* species in Himalayas and their uses with special reference to Uttaranchal. *Ethnobotany* 18: 53–66.
- Neshati F, Fritsch RM (2009) Seed characters and testa sculptures of some Iranian *Allium* L. species (Alliaceae). *Feddes Repertorium* 120(5–6): 322–332. <https://doi.org/10.1002/fedr.200911112>
- Nguyen NH, Driscoll HE, Specht CD (2008) A molecular phylogeny of the wild onions (*Allium*; Alliaceae) with a focus on the western North American center of diversity. *Molecular Phylogenetics and Evolution* 47(3): 1157–1172. <https://doi.org/10.1016/j.ympev.2007.12.006>
- Pandey A, Pandey R, Negi KS, Radhamani J (2008) Realizing value of genetic resources of *Allium* in India. *Genetic Resources and Crop Evolution* 55(7): 985–994. <https://doi.org/10.1007/s10722-008-9305-2>
- Pandey A, Pradheep K, Negi KS (2017) Onion and related taxa: ecogeographical distribution and genetic resources in Indian subcontinent. In: Ansari AA, Gill I, Singh, Abbas I, Naeem M (Eds) *Plant biodiversity: monitoring, assessment and conservation*. Wallingford, Oxfordshire, Boston MA, CABI International, 429–442. <https://doi.org/10.1079/9781780646947.0429>
- Pandey A, Malav PK, Rai KM, Ahlawat SP (2021) ‘Neodomesticates’ of the Himalayan allium spices (*Allium* species) in Uttarakhand, India and studies on morphology and eco-geography. *Genetic Resources and Crop Evolution* 68(5): 2167–2179. <https://doi.org/10.1007/s10722-021-01164-x>
- Seregin AP, Anačkov G, Friesen N (2015) Molecular and morphological revision of the *Allium saxatile* group (Amaryllidaceae): Geographical isolation as the driving force of underestimated speciation. *Botanical Journal of the Linnean Society* 178(1): 67–101. <https://doi.org/10.1111/boj.12269>
- Shah NC (2014) Status of cultivated and wild *Allium* species in India: A review. *Sciences et Techniques (Paris)* 1(9): 28–36.

- Sinitsyna TA, Herden T, Friesen N (2016) Dated phylogeny and biogeography of the Eurasian *Allium* section *Rhizirideum* (Amaryllidaceae). *Plant Systematics and Evolution* 302(9): 1311–1328. <https://doi.org/10.1007/s00606-016-1333-3>
- White TJ, Bruns T, Lee S, Taylor JW (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) *PCR protocols: a guide to methods and applications*. Academic Press Inc, New York, 315–322. <https://doi.org/10.1016/B978-0-12-372180-8.50042-1>
- Xie DF, Yu HX, Price M, Xie C, Deng YQ, Chen JP, Yu Y, Zhou SD, He XJ (2019) Phylogeny of Chinese *Allium* species in section *Daghestanica* and adaptive evolution of *Allium* (Amaryllidaceae, Allioideae) species revealed by the chloroplast complete genome. *Frontiers in Plant Science* 10: e460. <https://doi.org/10.3389/fpls.2019.00460>
- Xu JM, Kamelin RV (2000) *Allium* L. In: Wu ZY, Raven PH (Eds) *Flora of China*, vol. 24. Flagellariaceae through Marantaceae. Science Press, Beijing and Missouri Botanical Garden Press, St. Louis, 165–202.